

# Facile catalyzed acylation of heteroatoms using BiCl<sub>3</sub> generated in situ from the procatalyst BiOCl and acetyl chloride

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**Abstract**—Acylation of a variety of alcohols, phenols, aliphatic and aromatic amines, a thiol and a thiophenol proceeds efficiently using BiCl<sub>3</sub> generated in situ from the procatalyst BiOCl and acetyl chloride in a solvent or under solventless conditions, furnishing the corresponding acylated derivatives in very good to excellent yields.

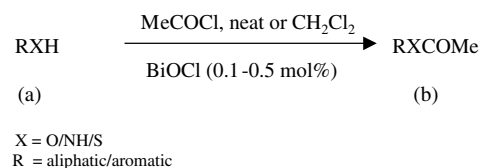
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Protection of heteroatoms by acylation is a very frequently employed organic transformation because of the mild deprotection methods available.<sup>1</sup> Acylation of hydroxy groups can be effected by carboxylic acids or derivatives like anhydrides,<sup>1</sup> acid chlorides,<sup>1</sup> acyl imidazoles<sup>2a</sup> or acyl ureas<sup>2b</sup> in the presence of bases such as Et<sub>3</sub>N, pyridine, DMAP,<sup>3a</sup> Bu<sub>3</sub>P<sup>3b</sup> or Lewis acids such as MgBr<sub>2</sub>,<sup>4a</sup> ZnCl<sub>2</sub>,<sup>4b</sup> CoCl<sub>2</sub>,<sup>4c</sup> TaCl<sub>5</sub>,<sup>4d</sup> InCl<sub>3</sub>,<sup>4e</sup> ZrCl<sub>4</sub><sup>4f</sup> and RuCl<sub>3</sub>.<sup>4g</sup> Some recent reports include metal triflates,<sup>5</sup> perchlorates<sup>6</sup> or supported reagents like HBF<sub>4</sub>–SiO<sub>2</sub>,<sup>7a</sup> HClO<sub>4</sub>–SiO<sub>2</sub>,<sup>7b</sup> KF–Al<sub>2</sub>O<sub>3</sub>,<sup>7c</sup> Nafion-H<sup>7d</sup> or yttria–zirconia,<sup>7e</sup> clay<sup>7f</sup> and zeolite HSZ-360.<sup>7g</sup> Iodine,<sup>8</sup> ionic liquids,<sup>9</sup> twisted amides<sup>10</sup> and *Pseudomonas capacia* PS lipase absorbed on Celite<sup>11</sup> have also been employed for acetylation of alcohols and phenols. However, the reported methods are not always devoid of disadvantages in terms of yields, chemoselectivity, moisture sensitivity, load and cost of the catalysts. Sometimes, the reagents are highly toxic, flammable or explosive. Thus, the search for newer mild and efficient methodologies for acylation continues.

Very recently, Bi(III) salts such as Bi(OTf)<sub>3</sub><sup>5c</sup> and BiOClO<sub>4</sub><sup>6c</sup> have been reported to effect acylation of heteroatoms with acid anhydrides. Bi(III) salts have served as Lewis acid catalysts for various other organic reactions<sup>12</sup> and are particularly important because of their low toxicity<sup>13</sup> and low cost; amongst these, BiCl<sub>3</sub> has been widely studied. BiOCl is an oxy salt of Bi(III) whose Lewis acidity is very low<sup>6c</sup> but it can generate

BiCl<sub>3</sub> on reaction with an acyl chloride and thus can act as a procatalyst and has been utilized in Friedel–Crafts acylation reactions.<sup>14</sup> The toxicity level of BiOCl is very low<sup>13</sup> and it is highly stable and readily available. We report herein the efficacy of BiOCl, in catalytic amounts, for the in situ generation of BiCl<sub>3</sub> vis-à-vis acylation of different heteroatoms in reactions with acyl chlorides (Scheme 1 and Table 1).

At the outset, β-naphthol was chosen as a model substrate and the minimum load of procatalyst and the acylating reagent (acetyl chloride) was ascertained using varying amounts of these in different solvents. The best results were obtained on treatment of β-naphthol in the



**Scheme 1.**

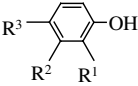
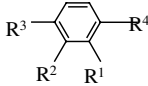
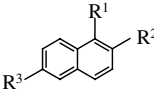
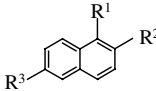
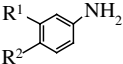
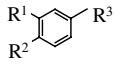


**Table 1.** Acetylation of β-naphthol based on BiOCl (0.5mol%) and AcCl (2equiv) in different solvents at room temperature

Entry	Solvent	Time	Yield (%)
1	CH <sub>2</sub> Cl <sub>2</sub>	45 min	94
2	CH <sub>3</sub> CN	1 h	91
3	THF	12 h	53
4	PhCH <sub>3</sub>	12 h	85

**Keywords:** Catalyzed; Acylation; Procatalyst; BiOCl; BiCl<sub>3</sub>.

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**Table 2.** Acylation of heteroatoms based on BiOCl–acyl chloride

Entry	Substrate (RXH) (a)	Method A <sup>a</sup> (CH <sub>2</sub> Cl <sub>2</sub> )		Method B <sup>b</sup> (neat)		Product <sup>c</sup> (RXAc) (b)
		Time	Yield (%)	Time	Yield (%)	
1	R = CH <sub>3</sub> (CH <sub>2</sub> ) <sub>3</sub> –	10 min	92	2 min	95	R = CH <sub>3</sub> (CH <sub>2</sub> ) <sub>3</sub> –
2	R = CH <sub>3</sub> (CH <sub>2</sub> ) <sub>5</sub> –	10 min	93	0.5 min	97	R = CH <sub>3</sub> (CH <sub>2</sub> ) <sub>5</sub> –
3	R = CH <sub>3</sub> (CH <sub>2</sub> ) <sub>7</sub> –	30 min	95	1 min	96	R = CH <sub>3</sub> (CH <sub>2</sub> ) <sub>7</sub> –
4	R = (CH <sub>3</sub> ) <sub>3</sub> C–	15 min	93	4 min	96	R = (CH <sub>3</sub> ) <sub>3</sub> C–
5	R = PhCH <sub>2</sub> –	5 min	98	0.5 min	97	R = PhCH <sub>2</sub> –
6	R = Ph(CH <sub>2</sub> ) <sub>2</sub> –	5 min	96	0.5 min	97	R = Ph(CH <sub>2</sub> ) <sub>2</sub> –
7	Mannitol	1.5 d	94	4 h min	98	Mannitol hexaacetate
8	Cyclohexanol	5 min	94	2 min	92	Cyclohexyl acetate
9	Benzoin	1.5 h	96	5 min	96	Benzoin acetate
10	Cholesterol	30 min	96	10 min	94	Cholesteryl acetate
11	Methyl α-D-Glcp	5 h	94	1.5 h	98	Methyl α-D-Glcp (OAc) <sub>4</sub>
12	(+)-Menthol	5 min	93	—	—	Menthyl acetate
13	Allyl alcohol	3 min	86	—	—	Allyl acetate
14	Propargyl alcohol	6 min	92	—	—	Propargyl acetate
						
15	R <sup>1</sup> = CO <sub>2</sub> Me, R <sup>2</sup> = R <sup>3</sup> = H	8 h	96	1.5 h	96	R <sup>1</sup> = CO <sub>2</sub> Me, R <sup>2</sup> = R <sup>3</sup> = H, R <sup>4</sup> = OAc
16	R <sup>1</sup> = R <sup>2</sup> = H, R <sup>3</sup> = Br	30 min	98	1 min	98	R <sup>1</sup> = R <sup>2</sup> = H, R <sup>3</sup> = Br, R <sup>4</sup> = OAc
17	R <sup>1</sup> = R <sup>3</sup> = H, R <sup>2</sup> = Me	2.5 h	90	5 min	98	R <sup>1</sup> = R <sup>3</sup> = H, R <sup>2</sup> = Me, R <sup>4</sup> = OAc
18	R <sup>1</sup> = OH, R <sup>2</sup> = R <sup>3</sup> = H	1 h	84	15 min	98	R <sup>1</sup> = OAc, R <sup>2</sup> = R <sup>3</sup> = H, R <sup>4</sup> = OAc
19	R <sup>1</sup> = R <sup>3</sup> = H, R <sup>2</sup> = OH	2.5 h	91	5 min	98	R <sup>1</sup> = R <sup>3</sup> = H, R <sup>2</sup> = OAc, R <sup>4</sup> = OAc
20	R <sup>1</sup> = R <sup>2</sup> = H, R <sup>3</sup> = OH	30 min	96	5 min	98	R <sup>1</sup> = R <sup>2</sup> = H, R <sup>3</sup> = OAc, R <sup>4</sup> = OAc
21	R <sup>1</sup> = NO <sub>2</sub> , R <sup>2</sup> = R <sup>3</sup> = H	6 h	49 <sup>d</sup>	2.5 h	96 <sup>h</sup>	R <sup>1</sup> = NO <sub>2</sub> , R <sup>2</sup> = R <sup>3</sup> = H, R <sup>4</sup> = OAc
22	R <sup>1</sup> = R <sup>3</sup> = H, R <sup>2</sup> = NO <sub>2</sub>	3 h	93	30 min	95	R <sup>1</sup> = R <sup>3</sup> = H, R <sup>2</sup> = NO <sub>2</sub> , R <sup>4</sup> = OAc
23	R <sup>1</sup> = R <sup>2</sup> = H, R <sup>3</sup> = NO <sub>2</sub>	5 h	95	30 min	98	R <sup>1</sup> = R <sup>2</sup> = H, R <sup>3</sup> = NO <sub>2</sub> , R <sup>4</sup> = OAc
24	R <sup>1</sup> = Ac, R <sup>2</sup> = R <sup>3</sup> = H	24 h	92	12 h	91	R <sup>1</sup> = Ac, R <sup>2</sup> = R <sup>3</sup> = H, R <sup>4</sup> = OAc
						
25	R <sup>1</sup> = R <sup>3</sup> = H, R <sup>2</sup> = OH	45 min	94	3 min	93	R <sup>1</sup> = R <sup>3</sup> = H, R <sup>2</sup> = OAc
26	R <sup>1</sup> = R <sup>3</sup> = H, R <sup>2</sup> = OH	30 min	95 <sup>f</sup>	3 min	94 <sup>f</sup>	R <sup>1</sup> = R <sup>3</sup> = H, R <sup>2</sup> = OAc
27	R <sup>1</sup> = R <sup>3</sup> = H, R <sup>2</sup> = OH	30 min	93 <sup>f,g</sup>	3 min	95 <sup>f,g</sup>	R <sup>1</sup> = R <sup>3</sup> = H, R <sup>2</sup> = OAc
28	R <sup>1</sup> = OH, R <sup>2</sup> = R <sup>3</sup> = H	12 h	95	5 min	94	R <sup>2</sup> = R <sup>3</sup> = H, R <sup>1</sup> = OAc
29	R <sup>1</sup> = H, R <sup>2</sup> = R <sup>3</sup> = OH	1 h	98	2 h	92	R <sup>2</sup> = R <sup>3</sup> = OAc, R <sup>1</sup> = H
30	R <sup>1</sup> = R <sup>3</sup> = H, R <sup>2</sup> = OH	19 h	70	5 h	93	R <sup>1</sup> = R <sup>3</sup> = H, R <sup>2</sup> = OBz
31	R <sup>1</sup> = OH, R <sup>2</sup> = R <sup>3</sup> = H	1.5 d	88	12 h	97	R <sup>1</sup> = OBz, R <sup>2</sup> = R <sup>3</sup> = H
32	R <sup>1</sup> = R <sup>3</sup> = H, R <sup>2</sup> = OH	12 h	95	—	—	R <sup>1</sup> = R <sup>3</sup> = H, R <sup>2</sup> = OOCCH <sub>2</sub> Ph
33	CH <sub>3</sub> (CH <sub>2</sub> ) <sub>7</sub> OH	1.5 d	94	6 h	91	CH <sub>3</sub> (CH <sub>2</sub> ) <sub>7</sub> OBz
						
34	R <sup>1</sup> = H, R <sup>2</sup> = Cl	10 min	96 <sup>e</sup>	10 min	95	R <sup>1</sup> = H, R <sup>2</sup> = Cl, R <sup>3</sup> = NHAc
35	R <sup>1</sup> = NO <sub>2</sub> , R <sup>2</sup> = H	5 min	94 <sup>e</sup>	—	—	R <sup>1</sup> = NO <sub>2</sub> , R <sup>2</sup> = H, R <sup>3</sup> = NHAc
36	R <sup>1</sup> = H, R <sup>2</sup> = NO <sub>2</sub>	5 min	94 <sup>e</sup>	3 min	93	R <sup>1</sup> = H, R <sup>2</sup> = NO <sub>2</sub> , R <sup>3</sup> = NHAc
37	R <sup>1</sup> = OH, R <sup>2</sup> = H	5.5 h	94 <sup>e</sup>	3 h	93	R <sup>1</sup> = OAc, R <sup>2</sup> = H, R <sup>3</sup> = NHAc
38	PhCH <sub>2</sub> NH <sub>2</sub>	30 min	98 <sup>e</sup>	—	—	PhCH <sub>2</sub> NHAc
39	CH <sub>3</sub> CH <sub>2</sub> SH	30 min	96	4 min	95	CH <sub>3</sub> CH <sub>2</sub> SAc
40		40 min	92	5 min	98	

<sup>a</sup> Using 0.5 mol% BiOCl.<sup>b</sup> Using 0.1 mol% BiOCl.<sup>c</sup> All products were characterized by IR, NMR and also by comparing the physical data with those of known compounds.<sup>d</sup> In refluxing (CH<sub>2</sub>Cl)<sub>2</sub>.<sup>e</sup> In CH<sub>3</sub>CN.<sup>f</sup> Scale-up (~30-fold) experiment.<sup>g</sup> With recovered BiOCl.<sup>h</sup> Under sonication.

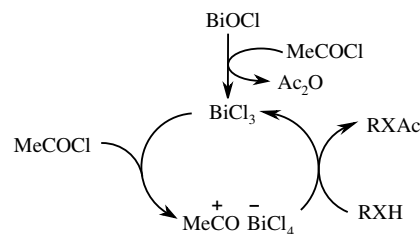
presence of 0.5 mol% BiOCl and acetyl chloride (2 equiv) in dichloromethane, which generated the  $\beta$ -naphthyl acetate in 45 min and in 94% yield (Table 1, entry 1; Table 2, entry 25). Yields in acetonitrile, tetrahydrofuran and toluene were 91%, 53% and 85%, respectively (Table 1; entries 2–4). Similar acetylation of  $\alpha$ -naphthol (entry 28), 2,6-dihydroxynaphthalene (entry 29) and other phenolic compounds containing electron withdrawing (entries 15, 21–24) and electron donating substituents (entries 16–20) on the aromatic ring proceeded satisfactorily in almost quantitative yields (Table 2). While acetylation of *o*-nitrophenol in refluxing dichloroethane was sluggish, complete conversion was accomplished in neat conditions after sonication for 2.5 h (Table 2, entry 21). Compounds containing primary (entries 1–3, 5,6), secondary (entries 7–12) and tertiary (entry 4) hydroxyl groups were easily acetylated in excellent yields (Table 2) under the present conditions without any competitive side reactions of the secondary and tertiary alcohols. Quantitative acetylation of *D*-mannitol and  $\alpha$ -*D*-glucopyranoside (entries 7 and 11, Table 2) was achieved with no competitive acetylation of the anomeric methoxy group<sup>15</sup> in case of the second substrate. The method is also applicable to the acetylation of chiral alcohols such as cholesterol and (+)-menthol (entries 10 and 12, Table 2) in very high yields and optical purity. Acetylation of allyl alcohol (entry 13) and propargyl alcohol (entry 14) also proceeded smoothly to their respective acetates in 86% and 92% yields (Table 2).

The scope of this simple acylation was further extended to the benzylation of  $\beta$ -naphthol,  $\alpha$ -naphthol and octanol (entries 30, 31 and 33, Table 2) with benzoyl chloride–BiOCl. However, a large excess of benzoyl chloride (~5 equiv) was necessary to effect complete benzylation. Similar acylation of  $\beta$ -naphthol with phenylacetyl chloride was also possible to give the corresponding ester with almost quantitative conversion (95%, Table 2, entry 32).

The generality of the present methodology was further established by the acetylation of amine and aniline derivatives. However, with these substrates, the best results were obtained in acetonitrile. Thus, each of *p*-chloroaniline (entry 34), *m*- and *p*-nitroanilines (entries 35 and 36), *m*-hydroxyaniline (entry 37) and benzylamine (entry 38) were readily acetylated in acetonitrile (Table 2). The general procedure of acetylation was equally applicable to ethanethiol and thiophenol (Table 2, entries 39 and 40), which yielded their respective acetylated products in 96% and 92% yields.

All the substrates could be acetylated under solvent-free conditions in excellent yields using even lower amounts of BiOCl (0.1 mol%) and in shorter reaction times (method B, Table 2), thus, making the present acylation procedure environmentally friendly.

The preparative efficacy of this procedure was verified by a scaling up (~30-fold) experiment with  $\beta$ -naphthol (Table 2, entry 26). Complete acetylation was possible with even 1.5 equiv of acetyl chloride. After aqueous work-up, BiOCl can be regenerated from the aqueous



**Figure 1.** In situ generation of BiCl<sub>3</sub> from BiOCl and its catalytic regeneration after acylation.

layer and can be reused with equal efficacy (Table 2, entry 27). A scale-up experiment (~30-fold) under neat conditions with BiOCl (0.1 mol%) and 1.5 equiv of acetyl chloride was also equally fruitful (yield 94%, method B, Table 2, entry 26).

As has already been noted, BiOCl in reaction with acyl chloride generates BiCl<sub>3</sub><sup>14</sup> in situ that in turn acts as a Lewis acid catalyst and activates the acylating reagent. After the acylation reaction, BiCl<sub>3</sub> is regenerated and thus the catalytic cycle (Fig. 1) is carried on.

The present method tolerates the presence of C=C, C≡C, CO<sub>2</sub>R, COR and NO<sub>2</sub> groups in the substrates and is thus a chemoselective procedure in the presence of the above functionalities. No Fries rearrangement products could be detected in the acylation of any of the phenolic substrates.

In conclusion, we have demonstrated that BiOCl, in reaction with an acyl chloride, is an efficient procatalyst for the in situ generation of BiCl<sub>3</sub> vis-à-vis acylation of alcohols, phenols, aliphatic and aromatic amines, a thiol and a thiophenol. The advantages of the present method are: BiOCl is an inexpensive, readily available and moisture stable procatalyst, the load of this reagent is quite low, and it has a very low toxicity level (LD<sub>50</sub> in rats, oral: 22 g/kg),<sup>13</sup> the reaction conditions are mild and simple, yields of the products are excellent in solution as well as in solventless conditions; moreover, BiOCl can be regenerated<sup>14</sup> and reused without any loss of its procatalytic activity. The conversion under solventless conditions along with the reusability, very low price and low toxicity level of the procatalyst make this method a more eco-friendly alternative to other existing methodologies and a suitable procedure for industrial application.

**General procedure for acetylation:** (a) Method A (in solvent)—To a mixture of substrate (1 equiv) and BiOCl (0.5 mol%) in CH<sub>2</sub>Cl<sub>2</sub> (or in CH<sub>3</sub>CN for amines, 4 ml) was added acetyl chloride (2 equiv) and the mixture was stirred at room temperature. After completion of the reaction (TLC) the mixture was diluted with CH<sub>2</sub>Cl<sub>2</sub>, then washed subsequently with brine (20 ml), saturated aq NaHCO<sub>3</sub> (2 × 15 ml) and H<sub>2</sub>O (20 ml). The organic layer was dried over Na<sub>2</sub>SO<sub>4</sub> and concentrated to dryness. The crude product was filtered through SiO<sub>2</sub> or crystallized before taking spectral data.

(b) Method B (under neat conditions)—To a mixture of substrate (1 equiv) and BiOCl (0.1 mol%) was added acetyl chloride (1.5 equiv) with stirring at room temperature. After completion of the reaction the mixture was processed as described in Method A.

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